

# Epilithic algae from an urban river preferentially use ammonium over nitrate

Eduardo Cejudo  · William D. Taylor · Sherry L. Schiff

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**Abstract** Epilithon is a taxonomically diverse assemblage of aquatic organisms which grow on rocks; this biological compartment is involved in several reactions that contribute to the dynamics of dissolved organic nitrogen in water. Using ceramic tiles colonized in an urban river, this experimental study assessed the relative importance of ammonium uptake by inhibiting nitrification blocking ammonium oxidation with acetylene, as well it tested the hypothesis that epilithon preferentially assimilated ammonium over nitrate. In our experiments, ammonium uptake by epilithon accounted for 46% – 100 % of the ammonium decrease in the water column, whereas nitrate uptake accounted for 0% – 11% of the nitrate decrease. Ammonium uptake rates ranged from 197 to 519  $\mu\text{mol N-NH}_4^+ \text{ m}^{-2} \text{ h}^{-1}$ , while nitrate uptake rates were from 47 to 85  $\mu\text{mol N-NO}_3^- \text{ m}^{-2} \text{ h}^{-1}$ . The rate of preferential assimilation (RPI) was between 1.15 and 1.26, indicating preference for ammonium over nitrate. The results of this research provide valuable information regarding the relative contribution of algal uptake relative to nitrification in epilithon from an urban river.

**Keywords** Epilithon · Uptake · Ammonium · Nitrate

## Introduction

Epilithon comprises organisms that grow on the surface of rocks in aquatic ecosystems. Epilithic biofilms in rivers play diverse roles in tightly coupled biogeochemical processes; for instance, photosynthesis and heterotrophic growth are often synergistic (Paerl and Pinckney 1996; Kuehn et al. 2014). Nutrient enrichment in rivers is commonly associated with an increase in biomass due to the response of the algal component (Stevenson et al. 2006; Elser et al. 2007; Kane et al. 2014); thus, the recovery of rivers impacted by nutrient point sources is largely dependent on the in-stream nutrient processing (Murdock et al. 2004; Ribot et al. 2012).

Epilithon assimilates ammonium ( $\text{NH}_4^+$ ) and nitrate ( $\text{NO}_3^-$ ) into biomass, whereas the mineralization of organic matter can regenerate  $\text{NH}_4^+$  (Wetzel 1964; Galloway 2003).  $\text{NH}_4^+$  is an energetically favorable nitrogen species because it requires less energy to be incorporated into organic nitrogen due to its reduced state (Taiz and Zeiger 2010). The expression  $\text{NH}_4^+$  preference is used to stress that, in spite of the availability of dissolved inorganic nitrogen, ammonium is taken up and incorporated into biomass at higher rates than nitrate. Preference for one species over the other is not simply inhibition or competition, it is rather a combination of processes affected by concentration and environmental conditions (Dortch 1990). Changes in environmental nitrogen concentrations do not necessarily become variations in uptake rates (Dortch 1990; Kemp and Dodds 2002; Ribot et al. 2013). N fluxes due to mineralization,  $\text{NH}_4^+$  oxidation or denitrification have been previously reported in epilithon (Duff et al. 1984; Binnerup et al. 1992; Kemp and Doods 2001; O'Brien et al. 2012). However, few papers have addressed autotrophic assimilation or the relative

Eduardo Cejudo (✉), William D. Taylor, Sherry L. Schiff  
Department of Earth Sciences, University of Waterloo. Waterloo, ON, Canada. N2L3G1  
e-mail: eduardo.cejudo@uwaterloo.ca

Eduardo Cejudo  
Department of Earth Sciences, University of Waterloo. Waterloo, ON, Canada. N2L3G1  
CONACYT – Centro de Investigación Científica de Yucatán A.C. Cancun, Quintana Roo, México. 77500.



contribution of bacterial versus algal uptake. For instance,  $\text{NH}_4^+$  assimilation by epilithon accounted for 9% – 23% of the total  $\text{NH}_4^+$  loss in streams (Hamilton et al. 2001; Peipoch et al. 2016). One way to disentangle the multiple nitrogen transformations that occur in epilithon is using chemical inhibitors. Several substances inhibit ammonium oxidation by biocidal activity or by causing toxicity to specific microbes that oxidize ammonium (Huber et al. 1977). The activity of the enzyme ammonia monooxygenase (AMO) is reduced or inhibited by direct binding with alternative substrates such as acetylene ( $\text{C}_2\text{H}_2$ ). Direct binding with an alternative substrate results into the irreversible inactivation of the enzyme and its recovery involves *de novo* protein synthesis (McCarty 1999). Acetylene has been previously used for simultaneously inhibiting both nitrification and denitrification (Knowles 1990; Teissier and Torre 2002). This type of chemical inhibition was resorted to in this research to distinguish autotrophic  $\text{NH}_4^+$  uptake from  $\text{NH}_4^+$  oxidation in epilithon; however, it does not account for heterotrophic uptake (Kirchman 1994). The objective of this research is to provide evidence regarding the importance of uptake and nitrification as pathways for  $\text{NH}_4^+$  loss in aquatic ecosystems and the preferential uptake of  $\text{NH}_4^+$  over  $\text{NO}_3^-$  by the autotrophic component in epilithon growing in the Grand River (southwestern Ontario, Canada) as an example of an urban river. Urban rivers receive large loads of N from wastewater treatment plants and information about nutrient processing is required (Gibson and Meyer 2007; Zhang et al. 2015). The Grand River (southwestern Ontario) is an example of an urban river impacted by economic activities. The land in the middle section of the watershed (where the epilithon for this research was grown) is used for intensive agriculture and urban development; thereby, high concentrations of nitrate and phosphorus are commonly measured owing to the extensive tile drainage and treated wastewater discharged from wastewater treatment plants (Grand River Conservation Authority 2006). Thus, understanding the preferential assimilation of one nitrogen species over the other will assist in assessing the impact of treated effluent into watercourses.

## Materials and methods

The middle section of the Grand River's watershed (southwestern Ontario) was used as an example of an urban river impacted by agriculture and urban development. The epilithon was grown on unglazed ceramic tiles, 25.8 cm<sup>2</sup> each. Sheets of tiles were incubated in uncovered plastic containers in the middle Grand River at a location 520 m downstream from Kitchener wastewater treatment plant's effluent (43.398° N, -80.4155° W) for 35 days (June 11<sup>th</sup> - July 17<sup>th</sup> 2013). Flow velocity and water depth varied over the growing period owing to fluctuating river discharges ( $40.8 \pm 27.2$  m<sup>3</sup>/s, HYDAT station 02GA048), result of summer rainfall and water released from two flow control structures in the upper Grand River (Conestogo and Shand Dams). Biomass was quantified from tiles that were not used in the experiments. The biomass from 16 tiles was brushed and scraped and the slurry obtained was dried for 24 h at 60°C, weighed, baked for 2 h at 550°C and then reweighed to ascertain ash-free dry weight (AFDW). Similar incubation experiments in the Grand River reported tile colonization by *Cladophora* spp, diatoms and unidentified green algae; however, macroscopic filamentous material was not observed in our samples (Barlow-Busch et al. 2006). Chlorophyll was measured from four tiles not used in the experiments. The slurry brushed off from the tiles was taken to a final volume of 500 mL with ultrapure water and kept in cold and dark conditions until analysis. The samples were filtered (0.5-0.7 bar) using pre-baked Whatman GF/F filters (0.7 μm). After filtration, the filters were folded and placed into 20-ml glass vials with 10 mL of 90% acetone, shaken gently and stored in darkness overnight (4°C). Before the analysis, samples were filtered to remove glass fibers from the liquid (0.45-μm Millipore syringe filter). The absorbance of both acetone blanks and extracts was measured at 750, 664, 647, and 630 nm and chlorophyll concentrations were calculated following Jeffrey and Humphrey (1975). Finally, chlorophyll *a* and ash-free dry mass (AFDM) were referred to as indicators of the community structure using the autotrophic index ( $\text{AI} = [\text{AFDM}/\text{Chl } a]$ ).

The tiles colonized with epilithon and used for “uptake” treatments were pre-incubated in a solution of 1%  $\text{C}_2\text{H}_2$  (v/v) overnight to inhibit nitrifying bacteria (following Herrmann et al. 2007). The chemical inhibition performed in this research assumes that nitrification was inhibited; hence,  $\text{NH}_4^+$  loss in such treatments was considered to represent autotrophic uptake, whereas treatments without  $\text{C}_2\text{H}_2$  accounted for “uptake + nitrification”. All treatments (in duplicate) had 300 mL of Grand-River water filtered through a 0.45-μm filter (Millipore filter membrane) in 400-mL HDPE containers (Starplex®). The treatments



**Table 1** Experimental design showing the nitrogen species present in each treatment (n=2) and the initial mass (C0 in  $\mu\text{mol N}$ ) for assessing uptake rates in epilithon. Inhibition refers to incubation with acetylene ( $\text{C}_2\text{H}_2$ , 1% v/v) prior to the experiment.

N specie(s)	C <sub>0</sub> [ $\mu\text{mol N}$ ]	Inhibition
$\text{NH}_4^+$	23	$\text{C}_2\text{H}_2$
	23	
	67	$\text{C}_2\text{H}_2$
	67	
	101	$\text{C}_2\text{H}_2$
	101	
$\text{NO}_3^-$	29	
	72	
	123	
$\text{NH}_4^+ + \text{NO}_3^-$	25 + 24	$\text{C}_2\text{H}_2$
	25 + 24	
	53 + 70	$\text{C}_2\text{H}_2$
	53 + 70	
	85 + 107	$\text{C}_2\text{H}_2$
	85 + 107	

were amended with three concentration levels of ammonium ( $(\text{NH}_4)_2\text{SO}_4$ , Fisher Scientific, CAS 7783-20-2) or nitrate ( $\text{KNO}_3$ , Fisher Scientific, CAS 7757-79-1) or both (Table 1). The initial concentration used in the experiments was similar to the annual average concentrations observed in the Grand River from 2010 to 2013 (0.1 to 0.8 mg N- $\text{NH}_4^+$  L<sup>-1</sup>; 3.0 to 4.0 mg N- $\text{NO}_3^-$  L<sup>-1</sup>); except for the heavy  $\text{NH}_4^+$  concentration treatment (4.7 mgN- $\text{NH}_4^+$  L<sup>-1</sup>) which was higher than any value ever observed in the river. All treatments started at the same time under the following conditions: 25° C, 24-hour light regime (630 lumens, incandescent bulbs), continuous stirring. Containers were lid-closed and each tile was elevated 2 cm from the bottom by means of a plastic support. Water subsamples from each experimental container were collected at 0, 0.5, 1, 2, 3, 6, 12, 24 and 48 h, filtered through a 0.45- $\mu\text{m}$  filter (0.45- $\mu\text{m}$  Millipore filter membrane) and stored in 5-mL HDPE vials for their later analyses.  $\text{NH}_4^+$  and  $\text{NO}_2^-$  were analyzed using colorimetric methods (blue indophenol and sulphanilamide + azo dye, respectively) with a Smartchem 200 Autoanalyzer (Westco, Brookfield CT; precision 5%, detection limit 0.05 mg N L<sup>-1</sup>).  $\text{NO}_3^-$  was analyzed with an Ion Chromatograph (Dionex ICS-90 Thermo Scientific, Sunnyvale CA; precision 0.07 mg N L<sup>-1</sup>, detection limit 0.05 mg N L<sup>-1</sup>). Table 1 shows the conditions that express initial N mass per experimental unit ( $\mu\text{mol N}$ ). Dissolved inorganic nitrogen (DIN) was calculated as the addition of  $\text{NO}_2^- + \text{NO}_3^- + \text{NH}_4^+$ . Since sampling continuously removed volume from all experimental units, mass removal due to volume withdrawal was accounted over time for accurate N mass per experimental unit.

$\text{NH}_3$  volatilization was estimated from the initial pH of the river's water amended with  $(\text{NH}_4)_2\text{SO}_4$  at pH = 7.6; this way, free ammonia was ascertained by means of equation 1

$$[\text{NH}_{3,\text{free}}] = \frac{[\text{NH}_4^+ \times 10^{\text{pH}}]}{(k_a/k_w) + 10^{\text{pH}}} \quad \text{Eq 1}$$

The estimated free  $\text{NH}_3$  was less than 2.3% of the total ammonium nitrogen (TAN) and was not considered in the DIN balance.

$\text{NH}_4^+$  and  $\text{NO}_3^-$  uptake rates ( $k_{\text{uptake}}$  in  $\mu\text{mol h}^{-1}$ ) were estimated as the slope of the linear regression of the N mass ( $\mu\text{mol N}$ ) by time (h) over the 48-hour duration of the experiment. Areal uptake rates were obtained for the colonized tiles ( $A = 25.8 \text{ cm}^2$ ) extrapolating to  $\mu\text{mol N m}^{-2} \text{ h}^{-1}$ . Finally, biomass-specific uptake rates ( $\mu\text{mol N g biomass}^{-1} \text{ h}^{-1}$ ) were ascertained by extrapolating the areal uptake rates measured in the tiles to the biomass that grew on them. Changes in biomass over the experiments (48 h) were not considered.



In order to quantify epilithon preference for  $\text{NH}_4^+$ , the rate of preferential incorporation (RPI, equation 2) was utilized following Dortch 1990, and Takahashi and Saijo 1981.

$$RPI_{\text{NH}_4^+} = \frac{P_{\text{NH}_4^+} / \Sigma \rho_N}{[\text{NH}_4^+] / [\Sigma N]} \quad \text{Eq 2}$$

Where  $P_{\text{NH}_4^+}$  is the ammonium uptake rate;  $\Sigma \rho_N$  is the sum of rates of all inorganic nitrogen species;  $[\text{NH}_4^+]$  ammonium concentration; and,  $[\Sigma N]$  is the total concentration of inorganic nitrogen. RPI  $\text{NH}_4^+$  values greater than 1.0 indicate preference for ammonium.

### Statistical analyses

Statistical analyses were run on JMP 5.1 (SAS Institute). Graphs were made in SPSS 13 (SPSS Inc.). Effect tests (F) and Least Squares Mean plots were produced to assess significant effects of the variables and their interactions. Post-hoc tests (Tukey Kramer HSD,  $\alpha=0.5$ ) were performed within treatments by N species to evaluate differences over time.

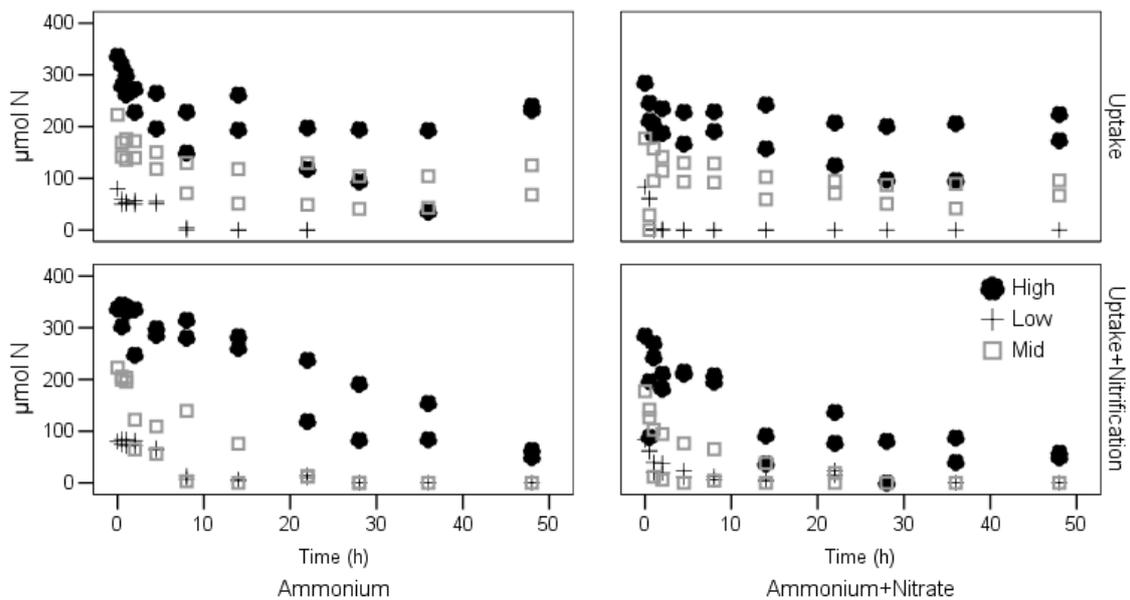
## Results

### Epilithic biomass measurements

The epilithon which grew on the tiles in the middle Grand River had 73.6 g biomass AFDW  $\text{m}^{-2}$  ( $\pm 37.2$  g  $\text{m}^{-2}$ ). Chlorophyll measurements were 42.4  $\pm 9.6$  mg Chl *a*  $\text{m}^{-2}$ , 8.7  $\pm 3.9$  mg Chl *b*  $\text{m}^{-2}$  and 3.6  $\pm 1.7$  mg Chl *c*  $\text{m}^{-2}$ . The Chl *a*: biomass ratio was  $\approx 0.0006$  mg Chl *a* (mg C AFDW) $^{-1}$  and the autotrophic index AI  $\approx 1700$ .

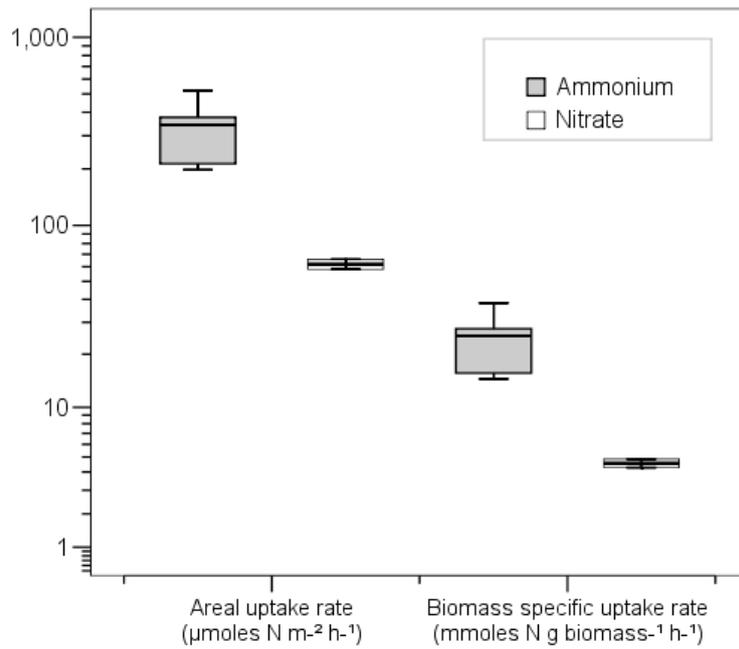
### $\text{NH}_4^+$ and $\text{NO}_3^-$ uptake

The change in  $\text{NH}_4^+$  ( $\text{NH}_4^+_{\text{initial}} - \text{NH}_4^+_{\text{final}}$ ) was different between treatments with and without inhibitor per concentration level (Tukey Kramer HSD,  $q = 2.47$ ,  $p = 0.01$ ,  $df = 10$ ). In the treatments without inhibitor

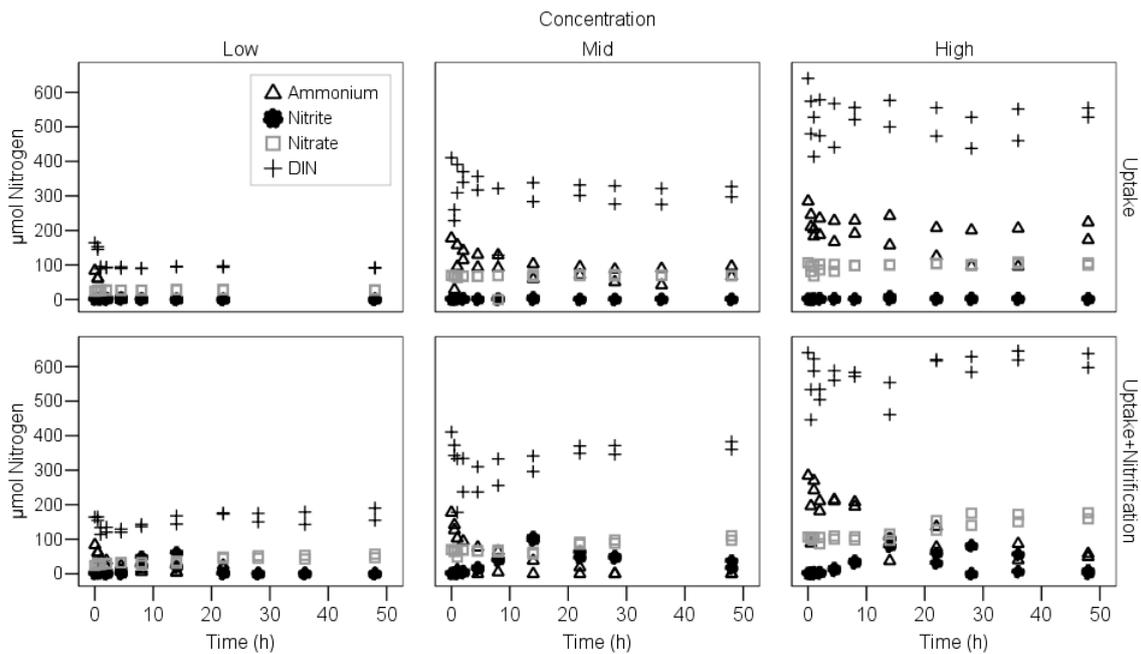


**Fig. 1** Ammonium decrease ( $\mu\text{mol N-NH}_4^+$ ) over time in treatments amended with ammonium only (*left*) and ammonium + nitrate (*right*). Uptake represents experimental units with tiles previously incubated with inhibitor (acetylene 1% v/v). Nitrogen concentrations (high, mid, low) as described in Table 1.





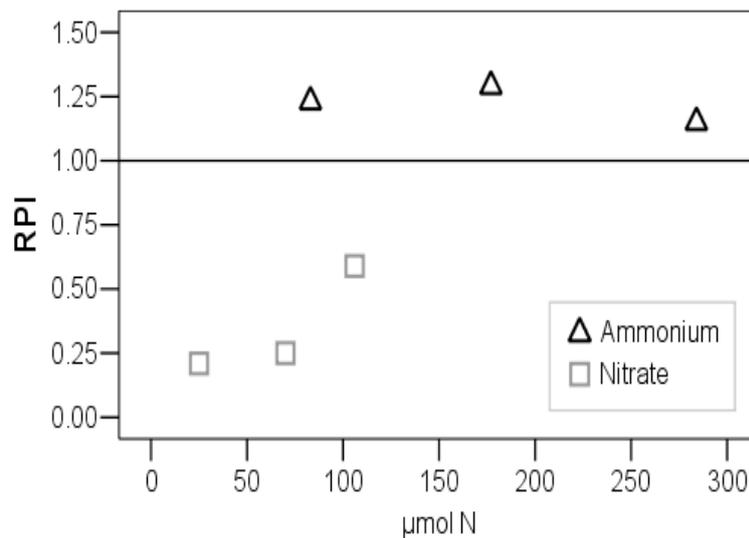
**Fig. 2** Estimated ammonium and nitrate areal uptake rates ( $\mu\text{mol N m}^{-2} \text{h}^{-1}$ ) and biomass specific uptake rates ( $\text{mmol N per g biomass}^{-1} \text{h}^{-1}$ ). NB: Y axis is in log scale.



**Fig. 3** Changes in ammonium, nitrite, nitrate and dissolved inorganic nitrogen (DIN) in treatments amended with ammonium + nitrate. Uptake represents experimental units with tiles previously incubated with inhibitor (acetylene 1% v/v). Nitrogen concentrations (high, mid, low) as described in Table 1

(*Uptake + Nitrification*) between 26% and 100% of the initial  $\text{NH}_4^+$  was removed from the water. The treatment with the highest concentration ( $> 101 \mu\text{mol N-NH}_4^+$ ) was the only one to display  $\text{NH}_4^+$  above the detection limit after 48 h. The treatments with inhibitor (*Uptake*) had a less steep decrease in  $\text{NH}_4^+$  (Fig. 1).  $\text{NH}_4^+$  decrease patterns were similar when both  $\text{NH}_4^+$  and  $\text{NO}_3^-$  were present in the same experimental unit. Only in the low concentration treatment was  $\text{NH}_4^+$  removed from the water column below detection limit after 48 h. Considering the change in  $\text{NH}_4^+$  from experimental units inhibited with  $\text{C}_2\text{H}_2$ , it was deemed that autotrophic uptake accounted for 46% and 56% decrease in the initial  $\text{NH}_4^+$  mass in the mid and high





**Fig. 4** Rate of preferential incorporation (RPI) of ammonium and nitrate in treatments amended with both nitrogen species at different initial N mass. Values above 1.0 represent  $\text{NH}_4^+$  preference.

concentration treatments, respectively. The change in  $\text{NO}_3^-$  was much less noticeable than the changes in  $\text{NH}_4^+$  in the mid and high concentration treatments, while it increased in the low concentration treatment despite the fact that  $\text{NH}_4^+$  was not added (not showed). The  $\text{NO}_3^-$  loss was 0 and 11% of the initial mass supplied for the mid and high concentration treatments, respectively.

The  $\text{NH}_4^+$  uptake rate ( $k_{\text{uptake}}$ ) in the treatments inhibited with  $\text{C}_2\text{H}_2$  was:  $0.97 \mu\text{mol N-NH}_4^+ \text{ h}^{-1}$  in the low concentration treatment ( $R^2 = 0.78$ ,  $\text{s.e.}_{\text{slope}} = 0.16$ );  $0.89 \mu\text{mol N-NH}_4^+ \text{ h}^{-1}$  in the mid concentration treatment ( $R^2 = 0.68$ ,  $\text{s.e.}_{\text{slope}} = 0.16$ ); and,  $1.34 \mu\text{mol N-NH}_4^+ \text{ h}^{-1}$  in the high concentration treatment ( $R^2 = 0.65$ ;  $\text{s.e.}_{\text{slope}} = 0.23$ ). The areal uptake rate in these experiments was between  $197.7$  and  $519.4 \mu\text{mol N-NH}_4^+ \text{ m}^{-2} \text{ h}^{-1}$  (Fig. 2).

$\text{NO}_3^-$  uptake rates ( $\text{NO}_3^- k_{\text{uptake}}$ ) were:  $0.15 \mu\text{mol N-NO}_3^- \text{ h}^{-1}$  in the mid concentration treatment ( $R^2 = 0.55$ ,  $\text{s.e.}_{\text{slope}} = 0.05$ ); and,  $0.17 \mu\text{mol N-NO}_3^- \text{ h}^{-1}$  in the high concentration treatment ( $R^2 = 0.41$ ,  $\text{s.e.}_{\text{slope}} = 0.03$ ).  $\text{NO}_3^-$  areal uptake was calculated from  $58$  to  $65 \mu\text{mol N-NO}_3^- \text{ m}^{-2} \text{ h}^{-1}$ .

Biomass specific uptake rates were calculated extrapolating the measured areal uptake rates to the biomass growth on tiles ( $73.6 \text{ g biomass AFDW m}^{-2}$ ). Thus, the estimated biomass specific uptake was  $24.4 \pm 8.7 \text{ mmol of N-NH}_4^+ \text{ g biomass}^{-1} \text{ h}^{-1}$  and  $4.6 \pm 0.3 \text{ mmol of N-NO}_3^- \text{ g biomass}^{-1} \text{ h}^{-1}$  (Fig. 2).

#### Interaction of $\text{NO}_3^-$ and $\text{NH}_4^+$

The third treatment assessed the dynamic of  $\text{NH}_4^+$  and  $\text{NO}_3^-$  together in the same experimental unit.  $\text{NH}_4^+$  decrease and  $\text{NO}_3^-$  increase were different between the concentrations assessed (Tukey Kramer HSD,  $q = 2.35$ ,  $p = 0.05$ ,  $\text{df} = 14$ ). The *Uptake + Nitrification* treatments showed the expected trend of  $\text{NH}_4^+$  decrease and later  $\text{NO}_3^-$  increase due to assimilatory and dissimilatory ammonium oxidation. The  $\text{NO}_2^-$  measured with  $\text{C}_2\text{H}_2$  inhibition was only 6% of the  $\text{NO}_2^-$  measured without  $\text{C}_2\text{H}_2$  (Fig. 3), providing information that nitrification was greatly reduced. The final DIN was always lower than the initial DIN between treatments; however, there was a less steep decrease in DIN in the treatments with inhibitor, which suggests that little N disappeared from the water column. The  $\text{NH}_4^+$  and  $\text{NO}_3^-$  uptake rates from the treatments, without inhibitor when the two species were present at the same time in our experimental units, were used to evaluate the rate of preferential incorporation (RPI), which showed that  $\text{NH}_4^+$  was preferred over  $\text{NO}_3^-$  ( $\text{RPI} > 1.0$ ; Fig. 4).

## Discussion

Preferential  $\text{NH}_4^+$  assimilation has been reported in submerged macrophytes (Schuurkes et al. 1986; Hood et al. 2014; Volkmann et al. 2016). The epilithic community from the middle Grand River grown on ceramic



tiles preferentially took up  $\text{NH}_4^+$  over  $\text{NO}_3^-$ , as indicated by the  $\text{NH}_4^+$  uptake rate, greater than the  $\text{NO}_3^-$  uptake rate, and from the RPI calculated when both N species were present. The observed pattern of  $\text{NH}_4^+$  decrease in the *Uptake + Nitrification* treatments was similar to the pattern observed for aerobic ammonia oxidizers in water (Sonthiphand et al. 2013) and sediments (Henriksen 1980). The  $\text{NH}_4^+$  removal due to uptake and nitrification in our experiments was higher than in other studies, a  $\text{NH}_4^+$  removal from 26% even up to 100% of the initial mass in each experimental unit was measured. Previous reports stated that the total contribution of epilithon to  $\text{NH}_4^+$  removal oscillated from 18% in a river that received treated wastewater (Ogura et al. 2009) to as much as 66% of the total N removal in grasslands streams (Simon et al. 2007). The  $\text{NH}_4^+$  and  $\text{NO}_3^-$  uptake rates measured in this research were higher than previously reported rates at lower dissolved inorganic nitrogen concentrations (Biggs 1990; Tank et al. 2000; Mulholland et al. 2004). We consider that the high concentration used in our experiments may have turned into higher experimental rates, as the biomass had a larger amount of dissolved nitrogen available per unit area, though it may also be due to the greater biomass that grew on the tiles, compared with other aquatic systems (Teissier et al. 2007).

*Chlorella* sp cultures showed a one-day delay in  $\text{NO}_3^-$  uptake when  $\text{NO}_3^-$  and  $\text{NH}_4^+$  were added simultaneously (Malerba et al. 2015).  $\text{NO}_3^-$  uptake in our experiments may have also been delayed and longer incubation periods might have produced different outcomes. The uptake of  $\text{NO}_3^-$  by epilithon in temperate forested streams has been reported from undetectable (Hamilton et al. 2001) to as much as  $850 \mu\text{mol N-NO}_3^- \text{ g biomass}^{-1} \text{ h}^{-1}$  (Triska et al. 1985). In our experiments,  $\text{NO}_3^-$  uptake by epilithon was small ( $< 25 \mu\text{mol N-NO}_3^- \text{ m}^{-2} \text{ h}^{-1}$ ) or non-measurable, and it was only observed at experimental conditions higher than  $72 \mu\text{mol N}$  ( $3.3 \text{ mg N-NO}_3^- \text{ L}^{-1}$ ). Epilithon uptake rates can be low compared with uptake rates of macrophytes on an areal basis due to the large biomass of macrophytes and the associated periphyton (Sand-Jensen and Borum 1991). Nonetheless, estimated gross N uptake by macrophytes with attached periphyton in different sections of the Grand River was similar to our results, from 3 to  $7.1 \mu\text{mol N per g macrophyte}^{-1} \text{ h}^{-1}$  (Hood 2012).

The chemical inhibition used in these experiments allowed us to distinguish the dissolved inorganic nitrogen dynamics and uptake between treatments distinguishing autotrophic *uptake* from *Uptake + Nitrification* in epilithon growing in an urban river. In our experiments, the calculated RPI supports the hypothesis that  $\text{NH}_4^+$  was preferred over  $\text{NO}_3^-$ . Preference for  $\text{NH}_4^+$  might be common because the reduced state of this N species confers an energetic advantage, as it is readily available for organic N synthesis (Lam et al. 1996). Pulses of  $^{15}\text{N}$  in the green algae *Enteromorpha intestinalis* showed also that more  $\text{NH}_4^+$  was removed and assimilated into tissue than  $\text{NO}_3^-$  (Cohen and Fong 2004). The observed low  $\text{NO}_3^-$  uptake rates could have been the result of the experimental conditions or else, the epilithon that colonized the tiles was not very effective removing nitrate. Given that uptake rates were measured as  $\text{NO}_3^-$  removed from the water column, it is also possible that  $\text{NO}_3^-$  increased in the experimental chambers as a result of organic matter mineralization (Böckelmann et al. 2000; Araya et al. 2003; Teissier et al. 2007), which was neither expected nor controlled in this research. The fact that some experimental units of the same treatment showed different uptake patterns suggested that there were differences in N cycling at biofilm scale, regardless of the same experimental treatment to all tiles.

In these experiments, the  $\text{NO}_3^-/\text{NH}_4^+$  mass ratio was similar across treatments (low=0.96; mid=1.3; and, high=1.25) and similar to the field conditions observed in the middle Grand River ( $\text{NO}_3^-/\text{NH}_4^+$  between 1 and 3 during the summers from 2010 to 2012). The biomass uptake rates reported in this research represent biofilm that grows in a river impacted by agricultural runoff and sewage discharges (Grand River Conservation Authority 2006).

Additional factors such as grazing (Hillebrand and Kahlert 2001), sediment abrasion (Francoeur and Biggs 2006) and the availability of other nutrients can modify the outcomes hereby reported. For instance, the large surface area of the tiles relative to the small volume of the experimental units can be an important factor when assessing nutrient assimilation rates (Nielsen and Jensen 1990). Photoinhibition (Merbt et al. 2017) and competitive exclusion of ammonia oxidizing bacteria by microalgae (Risgaard-Petersen et al. 2004) have been previously reported in experimental conditions. Regarding nutrient availability, the dissolved organic nitrogen (DON) was not measured; however, previously collected water samples in the middle Grand River downstream from the Kitchener wastewater treatment plant's effluent had an average DON of  $1.2 (\pm 0.6) \text{ mg N L}^{-1}$  (2011-2012, unpublished data), comprising between 12% and 25% of the total nitrogen measured in that reach of the river. It is possible that DON mineralization was an additional  $\text{NH}_4^+$



source. Organic matter mineralization was not expected to be an important factor during the experiment due to the abundant nutrient supply; however, there was an increase in  $\text{NO}_3^-$  at the low concentration treatments with  $\text{NO}_3^-$  as the only nitrogen source, possibly a result of internal  $\text{NH}_4^+$  regeneration. Finally, the biomass measured on the tiles was probably different from the biomass existent in the river throughout the year and such difference might have produced a different community composition. Previous studies measured 14 – 303 mg *chl a*  $\text{m}^{-2}$  (Teissier et al. 2007). Our results showed a more constrained range of chlorophyll ( $42.4 \pm 9.6$  mg *chl a*  $\text{m}^{-2}$ ); probably the result of the duration of the incubation and the relatively homogeneous community that grew on the tiles.

### Algal vs bacterial uptake

For the purposes of this section, uptake is understood as the introduction or translocation of a nutrient into cells, measured as the removal of the nutrient from the water column. Coupled assimilatory and dissimilatory  $\text{NH}_4^+$  removal have been amply recognized as important processes that transform and remove N from aquatic ecosystems (Sprent 1987; Bitton 2011); nevertheless, there is little information available on the relative contribution of algal uptake and assimilation compared with microbial oxidation. In this research, we generate knowledge on autotrophic uptake rates measured in an experiment with chemical inhibition estimated as  $>200 \mu\text{mol N-NH}_4^+ \text{m}^{-2} \text{h}^{-1}$  and  $>40 \mu\text{mol N-NO}_3^- \text{m}^{-2} \text{h}^{-1}$  for the epilithic community of an urban river. We consider these results valuable information regarding N removal from the water column with an experimental approach to algal uptake relative to bacterial ammonia oxidation. Reported  $\text{NH}_4^+$  uptake rates varied from 14.6 to  $50 \mu\text{mol N m}^{-2} \text{h}^{-1}$  and  $\text{NH}_4^+$  oxidation at a rate of  $1.16 \mu\text{mol N m}^{-2} \text{h}^{-1}$  (Peipoch et al. 2016). In our experiments, inhibition of dissimilatory  $\text{NH}_4^+$  oxidation highlighted the relevance of algal uptake, given that  $\text{C}_2\text{H}_2$  has been previously used for simultaneously inhibiting nitrification and denitrification (Teissier and Torre 2002). In like manner, a part of the  $\text{NO}_3^-$  decrease measured in our experiments is assumed to be algal uptake, though denitrification was not measured and its magnitude is unknown. The small *Chl a*: biomass ratio and the elevated autotrophic index suggest that the epilithic community had a large heterotrophic component or abundant organic detritus and materials other than biomass (Weitzel 1979; Biggs 1988; Cloern et al. 1995). Thus, it cannot be ruled out that the inhibited treatments also estimated assimilatory uptake by heterotrophic organisms instead of algal uptake alone.

The conceptual model of epilithic growth and maturity presented by Teissier et al. (2007) implies a tipping point where the biomass drives the balance of N removal. Assuming that the epilithic biomass grew on the tiles for these experiments was young, thin and actively growing, we can expect that N is taken up from the water column at rates exceeding mineralization and transient storage. This way, there were high uptake rates, low mineralization and reduced denitrification given that the biomass is thin enough and there is no limited  $\text{O}_2$  exchange within the biofilm and with the water column.

### Conclusions

This study showed that algal uptake was measured by chemical inhibition of nitrification using  $\text{C}_2\text{H}_2$ , and that  $\text{NH}_4^+$  was preferred over  $\text{NO}_3^-$  by the epilithic algae growth on ceramic tiles, as indicated by the RPI  $>1.0$ . Based on the rates obtained in these experiments, epilithon from the middle section of the Grand River assimilated  $\text{NH}_4^+$  in controlled conditions at rates of 197.7 to  $519.4 \mu\text{mol N-NH}_4^+ \text{m}^{-2} \text{h}^{-1}$ .  $\text{NO}_3^-$  assimilation rates were lower, from 58 to  $65 \text{N-NO}_3^- \text{m}^{-2} \text{h}^{-1}$  and sometimes  $\text{NO}_3^-$  was produced instead of being consumed. The estimated biomass specific uptake rates were  $24.4 \pm 8.7 \text{mmol of N-NH}_4^+ \text{g biomass}^{-1} \text{h}^{-1}$  and  $4.6 \pm 0.3 \text{mmol of N-NO}_3^- \text{g biomass}^{-1} \text{h}^{-1}$ . The uptake rates measured in these experiments provide valuable information on the contribution of algal uptake compared with nitrification in epilithic biofilms from an urban river.

**Competing interests** The authors declare they have no competing interests.

**Authors' contributions** EC designed and performed the experiment and prepared the manuscript. WDT and SLS contributed designing the experiment and analyzing the results. The authors read and approved the final manuscript.



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## References

- Araya R, Tani K, Takagi T, Tamaguchi N, Nasu M (2003) Bacterial activity and community composition in stream water and biofilm from an urban river determined by fluorescent in situ hybridization and DGGE analysis. *FEMS Microbiol Ecol* 43:113-119. doi: [10.1111/j.1574-6941.2003.tb01050.x](https://doi.org/10.1111/j.1574-6941.2003.tb01050.x)
- Barlow-Busch L, Baulch HM, Taylor WD (2006) Phosphate uptake by seston and epilithon in the Grand River, southern Ontario. *Aquat Sci* 68(2):181-192. <https://doi.org/10.1007/s00027-006-0806-9>
- Biggs BJF (1988) Artificial substrate exposure times for periphyton biomass estimates in rivers. *N Z J Mar Freshw Res* 22(4):507-515. <https://doi.org/10.1080/00288330.1988.9516321>
- Biggs BJF (1990) Use of relative specific growth rates of periphytic diatoms to assess enrichment of a stream. *N Z J Mar Freshw Res* 24(1):9-18. <https://doi.org/10.1080/00288330.1990.9516398>
- Binnerup SJ, Jensen K, Revsbech NP, Jensen MH, Sørensen J (1992) Denitrification, dissimilatory reduction of nitrate to ammonium, and nitrification in a bioturbated estuarine sediment as measured with  $^{15}N$  and microsensor techniques. *Appl Environ Microb* 58(1):303-313.
- Bitton G (2011) Wastewater microbiology. 4th ed. John Wiley and Sons. New Jersey USA.
- Böckelmann U, Manz W, Neu TR, Szewzyk U (2000) Characterization of the microbial community of lotic organic aggregates ('river snow') in the Elbe River of Germany by cultivation and molecular methods. *FEMS Microbiol Ecol* 33:157-170. <https://doi.org/10.1111/j.1574-6941.2000.tb00738.x>
- Cloern JE, Grenz C, Videgar-Lucas L (1995) An empirical model of the phytoplankton chlorophyll:carbon ratio—the conversion factor between productivity and growth rate. *Limnol Oceanogr* 40(7):1313-1321. <https://doi.org/10.4319/lo.1995.40.7.1313>
- Cohen RA, Fong P (2004) Nitrogen uptake and assimilation in *Enteromorpha intestinalis* (L.) Link (Chlorophyta): using  $^{15}N$  to determine preference during simultaneous pulses of nitrate and ammonium. *J Exp Mar Biol Ecol* 309(1):67-77. <https://doi.org/10.1016/j.jembe.2004.03.009>
- Dortch Q (1990) The interaction between ammonium and nitrate uptake in phytoplankton. *Mar Ecol Prog Ser* 61:183-201.
- Duff JH, Triska FJ, Oremland RS (1984) Denitrification associated with stream periphyton: chamber estimates from undisturbed communities. *J Environ Qual* 13(4):514-518. <https://doi.org/10.2134/jeq1984.00472425001300040002x>
- Elser JJ, Bracken ME, Cleland EE, Gruner DS, Harpole WS, Hillebrand H, Ngai JT, Seabloom EW, Shurin JB, Smith JE (2007) Global analysis of nitrogen and phosphorus limitation of primary producers in freshwater, marine and terrestrial ecosystems. *Ecol Lett* 10(12):1135-1142. <https://doi.org/10.1111/j.1461-0248.2007.01113.x>
- Francoeur SN, Biggs BJ (2006) Short-term effects of elevated velocity and sediment abrasion on benthic algal communities. *Hydrobiologia* 561(1):59-69. <https://doi.org/10.1007/s10750-005-1604-4>
- Galloway JN (2003) The Global Nitrogen Cycle. In: Schlesinger WH, Holland HD, Turekian K (eds) *Treatise on Geochemistry*, 8. Elsevier, pp 557-583
- Gibson CA and Meyer J L (2007). Nutrient uptake in a large urban river. *J Am Water Res Assoc* 43(3): 576-587.
- Grand River Conservation Authority (2006) Water quality in the Grand River: A summary of current conditions (2000-2004) and long term trends. Prepared by: Sandra Cooke Senior Water Quality Supervisor. Available at [https://www.grandriver.ca/en/our-watershed/resources/Documents/Water\\_Quality\\_Conditions\\_2006.pdf](https://www.grandriver.ca/en/our-watershed/resources/Documents/Water_Quality_Conditions_2006.pdf)
- Hamilton SK, Tank JL, Raikow DF, Wollheim WM, Peterson BJ, Webster JR (2001) Nitrogen uptake and transformation in a midwestern US stream: a stable isotope enrichment study. *Biogeochemistry* 54(3): 297-340. <https://doi.org/10.1023/A:1010635524108>
- Henriksen K (1980) Measurement of in-situ rates of nitrification in sediment. *Microbiol Ecol* 6:329-337. <https://doi.org/10.1007/BF02010495>
- Herrmann A, Witter E, Kätterer T (2007) Use of acetylene as a nitrification inhibitor to reduce biases in gross N transformation rates in a soil showing rapid disappearance of added ammonium. *Soil Biol Biochem* 39(9):2390-2400. <https://doi.org/10.1016/j.soilbio.2007.04.014>
- Hillebrand H, Kahlert M (2001). Effect of grazing and nutrient supply on periphyton biomass and nutrient stoichiometry in habitats of different productivity. *Limnol Oceanogr* 46(8), 1881-1898. <https://doi.org/10.4319/lo.2001.46.8.1881>
- Hood JLA (2012) The role of submersed macrophytes in river eutrophication and biogeochemical nutrient cycling. PhD dissertation, University of Waterloo, Canada.
- Hood JL, Taylor WD, Schiff SL (2014) Examining the fate of WWTP effluent nitrogen using  $\delta^{15}N-NH_4^+$ ,  $\delta^{15}N-NO_3^-$  and  $\delta^{15}N$  of submersed macrophytes. *Aquat Sci* 76(2):243-258. <https://doi.org/10.1007/s00027-013-0333-4>
- Huber DM, Warren HL, Nelson DW, Tsai CY (1977) Nitrification inhibitors—New tools for food production. *Bioscience* 27(8):523-529. <https://doi.org/10.2307/1297812>
- Jeffrey SW, Humphrey GF (1975) New spectrophotometric equations for determining chlorophylls a, b, c1 and c2 in higher plants, algae and natural phytoplankton. *Biochem Physiol Pfl* 167(2):191-194. [https://doi.org/10.1016/S0015-3796\(17\)30778-3](https://doi.org/10.1016/S0015-3796(17)30778-3)
- Kane DD, Conroy JD, Richards RP, Baker DB, Culver DA (2014) Re-eutrophication of Lake Erie: Correlations between tributary nutrient loads and phytoplankton biomass. *J Great Lakes Res* 40(3):496-501. <https://doi.org/10.1016/j.jglr.2014.04.004>
- Kemp MJ, Doods WK (2001) Centimeter-scale patterns in dissolved oxygen and nitrification rates in a prairie stream. *J N Am Benthol Soc* 20(3):347-357.
- Kemp MJ, Dodds WK (2002) The influence of ammonium, nitrate, and dissolved oxygen concentrations on uptake, nitrification, and denitrification rates associated with prairie stream substrata. *Limnol Oceanogr* 47(5):1380-1393. <https://doi.org/10.4319/lo.2002.47.5.1380>
- Kirchman DL (1994) The uptake of inorganic nutrients by heterotrophic bacteria. *Microb Ecol* 28(2):255-271. <https://doi.org/10.1007>



BF00166816

- Knowles R (1990) Acetylene inhibition technique: development, advantages, and potential problems. In: Denitrification in soil and sediment. Revsbech NP, Sorensen J (eds) Plenum Press, New York.
- Kuehn KA, Francoeur SN, Findlay RH, Neely RK (2014) Priming in the microbial landscape: periphytic algal stimulation of litter-associated microbial decomposers. *Ecology* 95:749-762. <https://doi.org/10.1890/13-0430.1>
- Lam HM, Coschigano KT, Oliveira IC, Melo-Oliveira R, Coruzzi GM (1996) The molecular-genetics of nitrogen assimilation into amino acids in higher plants. *Annu Rev Plant Phys* 47(1):569-593. <https://doi.org/10.1146/annurev.arplant.47.1.569>
- Malerba ME, Connolly SR, Heimann K (2015) An experimentally validated nitrate-ammonium-phytoplankton model including effects of starvation length and ammonium inhibition on nitrate uptake. *Ecol Model* 317:30-40. <https://doi.org/10.1016/j.ecolmodel.2015.08.024>
- McCarty GW (1999) Modes of action of nitrification inhibitors. *Biol Fert Soils* 29:1-9. <https://doi.org/10.1007/s003740050518>
- Merbt SN, Bernal S, Proia L, Martí E, Casamayor EO (2017) Photoinhibition on natural ammonia oxidizers biofilm populations and implications for nitrogen uptake in stream biofilms. *Limnol Oceanogr* 62(1):364-375. <https://doi.org/10.1002/lno.10436>
- Mulholland P, Valett HM, Webster JR, Thomas SA, Cooper LW, Hamilton SK, Peterson B (2004) Stream denitrification and total nitrate uptake rates measured using a field <sup>15</sup>N tracer addition approach. *Limnol Oceanogr* 49(3):809-820. <https://doi.org/10.4319/lno.2004.49.3.0809>
- Murdock J, Roelke D, Gelwick F (2004) Interactions between flow, periphyton, and nutrients in a heavily impacted urban stream: implications for stream restoration effectiveness. *Ecol Eng* 22(3):197-207. <https://doi.org/10.1016/j.ecoleng.2004.05.005>
- Nielsen SL, Jensen KS (1990) Allometric scaling of maximal photosynthetic growth rate to surface/volume ratio. *Limnol Oceanogr* 35(1):177-180.
- O'Brien JM, Hamilton SK, Podzikowski L, Ostrom N (2012) The fate of assimilated nitrogen in streams: an in situ benthic chamber study. *Freshw Biol* 57(6):1113-1125. <https://doi.org/10.1111/j.1365-2427.2012.02770.x>
- Ogura A, Takeda K, Nakatsubo T (2009) Periphyton contribution to nitrogen dynamics in the discharge from a wastewater treatment plant. *River Res App* 25:229-235.
- Paerl HW, Pinckney JL (1996) A mini-review of microbial consortia: their roles in aquatic production and biogeochemical cycling. *Microb Ecol* 31(3):225-247. <https://doi.org/10.1007/BF00171569>
- Peipoch M, Gacia E, Bastia E, Serra A, Proia L, Ribot M, Merbt SN, Martí E (2016) Small-scale heterogeneity of microbial N uptake in streams and its implications at the ecosystem level. *Ecology* 97:1329-1344. <https://doi.org/10.1890/15-1210.1>
- Ribot M, Martí E, von Schiller D, Sabater F, Daims H, Battin TJ (2012) Nitrogen processing and the role of epilithic biofilms downstream of a wastewater treatment plant. *Freshw Sci* 31(4):1057-1069. <https://doi.org/10.1899/11-161.1>
- Ribot M, Von Schiller D, Peipoch M, Sabater F, Grimm NB, Martí E (2013). Influence of nitrate and ammonium availability on uptake kinetics of stream biofilms. *Freshw Sci* 32(4):1155-1567. <https://doi.org/10.1899/12-209.1>
- Risgaard-Petersen N, Nicolaisen MH, Revsbech NP, Lomstein BA (2004) Competition between ammonia-oxidizing bacteria and benthic microalgae. *Appl Environ Microbiol* 70(9):5528-5537. <https://doi.org/10.1128/AEM.70.9.5528-5537.2004>
- Sand-Jensen K, Borum J (1991) Interactions among phytoplankton, periphyton and macrophytes in temperate freshwaters and estuaries. *Aquat Bot*, 41:137-175. [https://doi.org/10.1016/0304-3770\(91\)90042-4](https://doi.org/10.1016/0304-3770(91)90042-4)
- Schuurkes JA, Kok CJ, Den Hartog C (1986) Ammonium and nitrate uptake by aquatic plants from poorly buffered and acidified waters. *Aquat Bot* 24(2):131-146. [https://doi.org/10.1016/0304-3770\(86\)90093-8](https://doi.org/10.1016/0304-3770(86)90093-8)
- Simon KS, Niyogi DK, Frew RD, Townsend CR (2007) Nitrogen dynamics in grassland streams along a gradient of agricultural development. *Limnol Oceanogr* 52(3):1246-1257. <https://doi.org/10.4319/lno.2007.52.3.1246>
- Sonthiphand P, Cejudo E, Schiff SL, Neufeld JD (2013) Wastewater effluent impacts ammonia-oxidizing prokaryotes of the Grand River, Canada. *Appl Environ Microb* 79(23):7454-7465.
- Sprent J (1987) The ecology of the nitrogen cycle. Cambridge University Press, Great Britain.
- Stevenson RJ, Rier ST, Riseng CM, Schultz RE, Wiley MJ (2006) Comparing effects of nutrients on algal biomass in streams in two regions with different disturbance regimes and with applications for developing nutrient criteria. *Hydrobiologia* 561(1):149-165. [https://doi.org/10.1007/1-4020-5070-4\\_11](https://doi.org/10.1007/1-4020-5070-4_11)
- Taiz L, Zeiger E (2010). *Plant Physiology*. 5<sup>th</sup> ed. Sinauer Associates, Massachusetts U.S.A.
- Takahashi M, Saijo Y (1981) Nitrogen metabolism in Lake Kizaki, Japan I. Ammonium and nitrate uptake by phytoplankton [Algae]. *Arch Hydrobiol* 12(5):393-407.
- Tank JL, Meyer JL, Sanzone DM, Mulholland PJ, Webster JR, Peterson BJ, Leonard NE (2000). Analysis of nitrogen cycling in a forest stream during autumn using a <sup>15</sup>N tracer addition. *Limnol Oceanogr* 45:1013-1029
- Teissier S, Torre M (2002) Simultaneous assessment of nitrification and denitrification freshwater epilithic biofilms by acetylene block method. *Water Res* 36:3803-3811. [https://doi.org/10.1016/S0043-1354\(02\)00098-2](https://doi.org/10.1016/S0043-1354(02)00098-2)
- Teissier S, Torre M, Delmas F, Garabétian F (2007) Detailing biogeochemical N budgets in riverine epilithic biofilms. *J N Am Benthol Soc* 26(2):178-190. [https://doi.org/10.1899/0887-3593\(2007\)26\[178:DBNBIR\]2.0.CO;2](https://doi.org/10.1899/0887-3593(2007)26[178:DBNBIR]2.0.CO;2)
- Triska FJ, Kennedy VC, Avanzino RJ (1985) Uptake and regeneration of nitrate by epilithic communities in a nearly pristine lotic environment. Selected papers in the Hydrologic Sciences, USGS water-supply paper 2270 pp 91-98
- Volkman C, Halbedel S, Voss M, Schubert H (2016) The role of dissolved organic and inorganic nitrogen for growth of macrophytes in coastal waters of the Baltic Sea. *J Exp Mar Biol Ecol* 477:23-30. <https://doi.org/10.1016/j.jembe.2016.01.005>
- Weitzel RL (1979) Periphyton measurements and applications. In: Weitzel RL (ed) *Methods and measurements of periphyton communities: a review*. ASTM Special Technical Publication 690 United States, pp 3-33. <https://doi.org/10.1520/STP35061S>
- Wetzel RG (1964) A comparative study of the primary production of higher aquatic plants, periphyton, and phytoplankton in a large, shallow lake. *Int Rev Gesamten Hydrobiol* 49(1):1-61. *Methods and measurements of periphyton communities: a review* [doi: 10.1002/iroh.19640490102](https://doi.org/10.1002/iroh.19640490102)
- Zhang X, Wu Y, Gu B (2015) Urban rivers as hotspots of regional nitrogen pollution. *Environ Poll* 205:139-144. <https://doi.org/10.1016/j.envpol.2015.05.031>

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